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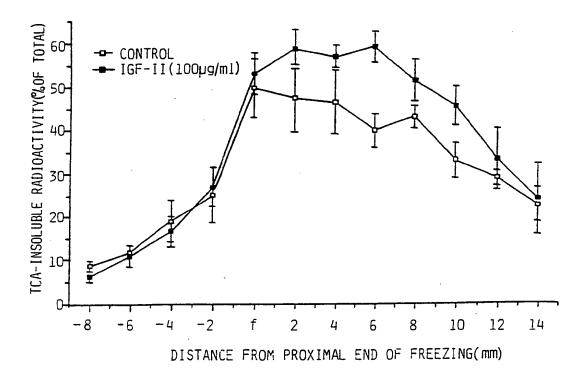
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(54) Title: NEW MEDICINAL USE



(57) Abstract

The use of IGF-II for preparing a medicament for nerve regeneration; and a method for medical treatment resulting in nerve regeneration, comprising the step of administering to patient in need of such treatment an effective amount of IGF-II.

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#### New medicinal use

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The present invention relates to a new medicinal use, more particularly the use of insulin-like growth factor II (IGF-II) for preparing a medicament for nerve repair and regeneration.

The nervous system of a vertebrate is subdivided into the central nervous system (CNS), which consists of the brain and the spinal cord, and the peripheral nervous system (PNS), which includes the cranial and spinal nerves and the peripheral components of the autonomic nervous system. The present invention involves new techniques enabling improved nerve regeneration and the invention is applicable to disorders or damages in the central nervous system (CNS) and the peripheral nervous system (PNS).

Insulin-like growth factor II (IGF-II) is structurally significantly different from IGF-I and little is known about its therapeutic usefulness. IGF-II is a single-chain peptide containing 67 amino acid residues. For further details concerning IGF-II see Eur. J. Biochem. 190, 445-462 (1990), René E. Humbel, Review Insulin-like Growth Factors I and II. IGF-II and IGF-I acts via different receptors. The receptor for IGF-II is a single chain receptor with a molecular weight of 260 kD and it is identical with the mannose-6-phosphate receptor (Morgan et al. Nature 329, 301-307 (1987); Kiess et al., J. Biol. Chem. 263, 9339-9344 (988). The IGF-II receptor has no homology with the IGF-I receptor which is dimeric and shows protein kinase activity. IGF-II may crossreact with the IGF-II receptor (Roth, R.A. Science 1988, 239, 1269-1271).

It is known that growth factors such as IGF-I, FGF and NGF promote neurite extension in cultured dorsal root ganglia cells or neuroblastoma cells [Recio-Pinto, E. t al. J.Neu-rosci. 6, 1211-1219 (1986); Morrison, R.S. et al. Proc.Natl-Acad.Sci. USA, 83, 7533-7541 (1986); Shelton, D.L., Proc-

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Natl.Acad.Sci, USA, 83, 2714-2718 (1986)]. However, it is to be noted that growth of axons in a regenerating peripheral nerve is a different process with respect to the influence of growth factors and drugs. Thus, regeneration of the sciatic nerve is unaffected by NGF [Rich, K.M., J. Comparative Neurology 230, 110-118 (1984)] and a variety of drugs used to induce neurite formation in culture. It also known that IGF-I stimulates nerve regeneration in vivo in rat. (Regeneration at the rat sciatic nerve, Dissertation Departement of Zoophysiology, University of Lund, Sweden, February 9, 1990, by Jacob Sjöberg).

#### In vitro data:

IGF-II has been demonstrated to stimulate neurite outgrowth and survival in vitro of cultured chick sympathetic and sensory neurons (Recio-Pinto E. et al., J.Neurosoci. 6, 1211-1219 (1986); Bothwell, J.Neurosci. Res 225-231 (1982)].

However, it could not be concluded from these results that IGF-II would also stimulate nerve regeneration in vivo, since eg. NGF is clearly effective in vitro but has no effect on nerve regneration in vivo. Furthermore, our results on nerve explants surprisingly showed that IGF-II affected the nerve-associated cells rather than the neurons themselves.

In connection with extensive research and experimentation it has now surprisingly been found that IGF-II effectively promotes nerve regeneration, albeit through a different mechanism than IGF-I, as will be shown below in the experimental section of this disclosure.

Administration of IGF-II in vivo showed that it significantly improved regneration of rat sciatic nerves subjected to a freeze lesion. Measurments of thymidine incorporation along the nerve following such administration suggested that IGF-II acted by stimulation of proliferation of nerve associated cells. This implication was further substantiated by experiments in culture which showed that IGF-II enhanced proliferation of nerve associated cells in cultured segments of the sciatic nerve. The latter experiments also demonstrated

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that IGF-II acts via different m chanisms than IGF-I which fails to affect proliferation in excis d s gments of the sciatic nerve.

The claims made in this application are primarily based on our finding that IGF-II stimulates regeneration of the sciatic nerve in adult rats. This stimulation appears to occur through activation of nerve associated cells. The findings, with respect to the effects on regneration and mechanism of action are new and surprising and affords new curative techniques for the treatment of damaged nervous tissue.

Accordingly, it is a main object of the present invention to provide new techniques for preparing medicaments effective in providing improved regeneration of nerves subjected to damage.

Another object of the invention is to provide for a medicament useful in the treatment of neuropathy.

A third object of the invention is to provide techniques for preparing a medicament for use in the treatment of degenerative neural disorders.

Yet another object is to provide techniques for preparing medicaments, wherein IGF-II is used in combination with IGF-I.

Accordingly, the invention resides in the use of IGF-II for preparing a medicament useful for nerve regeneration.
According to preferred aspects of the invention such medicament can be used in the treatment of neuropathy or, alternatively, in the treatment of degenerative neural disorders and survival of nervous tissue for transplantation.

The invention also resides in a method for medical treatment resulting in nerve regeneration, said method comprising the step of administering to a patient in need of treatment for nerve regeneration an effective amount of IGF-

In this method IGF-II can be used in combination with IGF-I. The mechanisms of action between IGF-I and IGF-II are different. IGF-I stimulates the neuronal cells and IGF-II.

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stimulates the nerve associated cells whereby a combination of the two gives an optimal effect of regeneration. The proportions between these two growth factors may vary between 1:10 to 10:1 based on weight.

The medicament used in accord with the present invention may thus in accordance with traditional pharmaceutical practice be formulated for use in human or veterinary medicine for therapeutic purposes. The compositions may include the active ingredient IGF-II in combination with a pharmaceutically or diganostically acceptable carrier, which may be solid, semisolid or liquid.

The compositions include those in a form adapted for topical application.

Suitable forms of the composition of this invention include tablets, capsules, sirups, suspensions, solutions, and forms suitable for injection or infusion. Such compositions may contain conventional pharmaceutically acceptable materials, such as diluents, binders, colours, flavours, preservatives, disintegrants and the like in accordance with conventional pharmaceutical practice in the manner well understood by those skilled in the art of formulating drugs.

Injectable or infusable compositions of IGF-II are particularly suitable as increased levels of IGF-II in the circulation can occur after administration by injection or infusion.

The invention will in the following be described by non-limiting examples with reference to the appended drawings, wherein:

Figure 1 is a diagram illustrating experiments involving effect of IGF-I at various concentrations on in vitro proliferation in the rat sciatic nerve;

Figure 2 is a corresponding diagram but using IGF-II instead of IGF-I;

Figure 3 shows a diagram on thymidine incorporation in freeze-injured rat sciatic nerve with and without treatment with IGF-II.

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Figure 4 shows a diagram on regeneration distance in freeze-injured rat sciatic nerve with and without treatment with IGF-II; and

Fig. 5 shows a corresponding diagram comparing the effect of regeneration using two different concentrations of IGF-II.

#### EXAMPLE 1

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Comparison between IGF-I and IGF-II in the rat sciatic nerve in vitro.

#### Animals

Female Sprague-Dawley rats weighing around 200 g were used. The rats were anaesthetized by an intraperitoneal injection of 0.3 ml of a 1:1.2 mixture of pentobarbital (60 mg/ml) saline (0.9%) and diazepam 5 mg/ml). Both sciatic nerves were then removed by dissection.

#### Culture

The dissected nerve was cut into 4 mm segments. These were transferred to plastic Petri dishes (Nunclon 3.5 cm) containing serum free RPMI 1640 medium without or with IGF-I or IGF-II at concentrations indicated in Fig. 1 and Fig. 2, respectively. Each dish was supplied with 3-4 nerve segments.

The preparations were incubated for 48 h at 37°C in a humidified atmosphere of 5% CO<sub>2</sub> in air.

#### Thymidine incorporation

Thymidine incorporation was measured essentially as describ d previously (J. Sjöberg & M. Kanje, Brain Research, 530 (1990) 167-169). The cultured segments were rinsed in rat Ringer solution and transferred to test tubes containing Ringer solution supplied with 50 µCi 3H-thymidine/ml (Amersham, specific activity 91 Ci/mmol). The tubes were incubated in a waterbath at 37°C for 2h. The segments were then washed in ice-cold Ringer solution and subsequently extracted in 10% tri-

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chloro acetic acid (TCA) at 4°C for a minimum of 2h. The segments were then washed in TCA and finally dissolved Soluene 350 over night. Radioactivity in the TCA-soluble and insoluble fractions was determined by liquid scintillation counting. Thymidine incorporation was expressed as TCA-insoluble radioactivity in percent of total radioactivity in the segment.

The results of the experiments on thymidine incorporation at varying concentrations of the respective growth factors are illustrated in Figs. 1 and 2.

Fig. 1 shows the results of IGF-I at varying concentrations and clearly shows that IGF-I at concentrations varying from 0.01 to 1 ng/ml has no significant effect on the proliferation in the sciatic nerve in vitro, while an inhibition was observed at 100 ng as compared to a control devoid of growth factor.

Fig. 2 illustrates a corresponding set of experiments using IGF-II and Fig. 2 clearly illustrates a significant increase of the <u>in vitro</u> proliferation in the interval between 0.1 and 1000 ng/ml. This is accordingly also an indication of the fact that IGF-II affects nerve growth by a different mechanism than does IGF-I.

### EXAMPLE 2

The stimulatory effect of IGF-II on regeneration of freeze-injured rat sciatic nerve.

### Animals

Female Sprague-Dawley rats (Alab, Sweden) weighing approximately 200 g were anesthetized by i.p. injection of 0.35 ml of a Nembutal (50 mg/ml)-Valium (5 mg/ml)-saline mixture (1:2:1 v/v). The sciatic nerve was exposed in the mid-thigh and frozen over a 15 mm segment with specially designed tweezers chilled in liquid nitrogen. Freezing was performed twice for 30 s with 2 min thawing in between. The proximal end of the frozen segment was labeled by attaching a 9-0 suture to the epineurium.

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An osmotic minipump (model 2001, Alzet, USA) filled with a modified Ringer solution (in mM): NaCL 139, KCl 2.4, MgSO<sub>4</sub> 2, NaH<sub>2</sub>PO<sub>4</sub> 0.6, Na<sub>2</sub>HPO<sub>4</sub> 3.25, pH 7.4 and different concentrations of IGF-1 (Kabi Pharmacia AB, Stockholm, Sweden) were implanted subcutaneously on the abdomen of the rat. The pump rate was 1.05 µl/h. From the pump a thin silicone catheter was drawn subcutaneously to the sciatic nerve. The catheter was sutured alongside the freeze-injured nerve with the catheter opening half-way down the freeze-injured segment (Fig.-1). The part of the catheter lying parallel to the nerve was perforated to increase the perfused area. Two animals were kept in each cage at a room temperature of 21°C and 52% humidity, and given water and food ad libitum.

#### EXAMPLE 3

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Thymidine incorporation

The animals were sacrificed 6 days after surgery and the sciatic nerves were removed. The nerves were desheathed and incubated for 2h in vials containing 50 µCi of [3H]thymidine (9i Ci/mmol) in 1 ml of modified Ringer solution at 37°C. After thorough washing with ice-cold Ringer solution the nerves were cut in 2 mm pieces and extracted for 2x30 min in 10% trichloroacetic acid (TCA). TCA-insoluble material was solubilized in 200 µl Soluene 350 (Packard). Radioactivity of TCA-soluble and insoluble fractions were determined by liquid scintillation counting. Thymidine incorporation was measured in the same way as in Example 1 esssentially according to the techniques described in the Sjöberg et al. Brain Research, 530 (1990) 167-169.

Fig. 3 shows a diagram illustrating the effect of IGF-II on the incorporation of thymidine along the nerve 6 days after freezing. Comparison is made with a control using no IGF-II. As can be seen from the diagram IGF-II perfusion results in a significant increase in proliferation compared to the control with no IGF-II perfusion. This <u>in vivo</u> experiment thus illustrates with clarity the usefulness of IGF-II in

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enhancing proliferation of regenerating nerve upon injury or other nerve cell disorder.

# EXAMPLE 4

Example 2 was repeated and regeneration distances after 6 days profusion of the freeze-injured nerve section with and without administration of IGF-II and at two different concentrations of IGF-II are shown in Fig. 4 and Fig. 5, respectively.

As seen from the results illustrated in Fig. 4 treatment with IGF-II at 100  $\mu g/ml$  results in a significant increase in regeneration distance as compared to the control.

Fig. 5 shows the dose response when using IGF-II at 25  $\mu g$  and 100  $\mu g/ml$ . In this figure is expressed as the difference in regeneration distance compared to the control. In Figs. 4 and 5 the asterisc means p< 0.05.

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#### CLAIMS

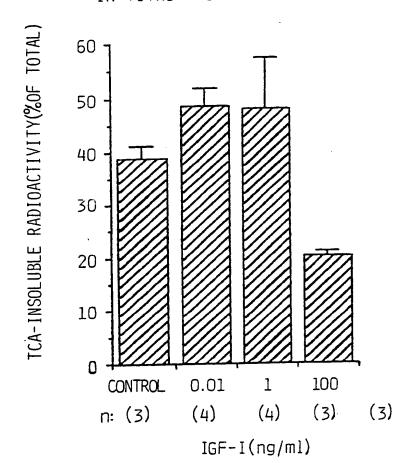
- 1. The use of IGF-II for preparing a medicament for nerve regeneration through activation of nerve associated cells.
- 2. The use according to claim 1 for preparing a medicament for use in the treatment of neuropathy.
- 3. The use according to claim 2 for preparing a medicament\_for use in the treatment of degenerative neural discorders.
- 4. The use according to any preceding claim, wherein IGF-II is used in combination with IGF-I.
  - 5. A method for medical treatment resulting in nerve regeneration, comprising the step of administering to a patient in need of such treatment an effective amount of IGF--II.
  - 6. A method according to claim 5 for the treatment of neuropathy.
  - 7. A method according to claim 5 for the treatment of degenerative neural disorders.
- 20 8. A method according to claim 5 as applied to using IGF-II for maintenance or survival of transplants.

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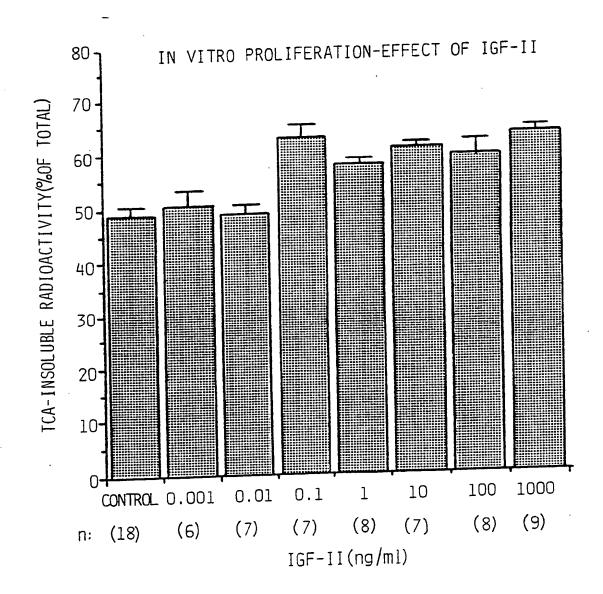
FIG.1

IN VITRO PROLIFERATION-EFFECT OF IGF-I



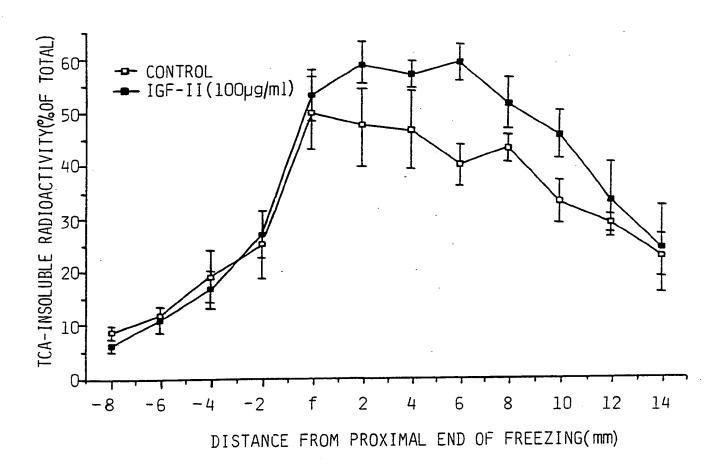
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FIG.2



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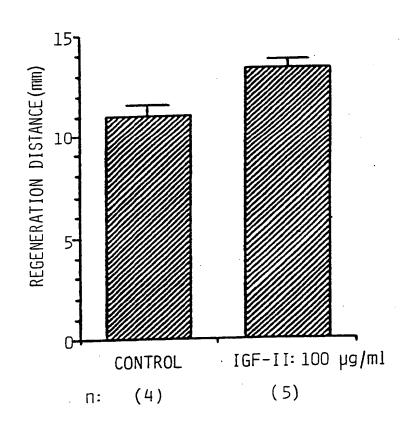
FIG.3



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FIG.4

REGENERATION DISTANCE OF FROZEN NERVE AFTER 6 DAYS

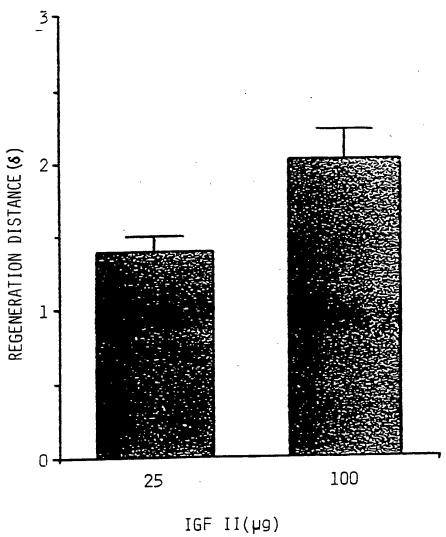


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FIG.5

EFFECTS OF IGF-II ON REGENERATION OF THE RAT SCIATIC NERVE



# INTERNATIONAL SEARCH REPORT

International Application No PCT/SE 92/00275

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III. DOCL	IMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)	Relevant to Claim No
Category "	Citation of Document, with indication, where appropriate, of the relevant passages	
x	Brain Research, Vol. 302, 1984 Recio-Pinto, E. et al.: "Effects of Insulin, Insulin-like Growth Factor-II and Nerve Growth Factor on Neurite Outgrowth in Cultured Human Neuroblastoma Cells", see page 323 - page 334	1-3
x	The Journal of Neuroscience, Vol. 6, No. 5, May 1986 Recio-Pinto, E. et al.: "Effects of Insulin, Insulin-like Growth Factor-II, and NerveGrowth Factor on Neurite Formation and Survival in Cultured Sympathetic and Sensory Neurons", see page 1211 - page 1219	1-3
x	WO, A1, 9102067 (MAX PLANCK-GESELLSCHAFT ZUR FÖRDERUNG DER WISSENCHAFTEN E.V.) 21 February 1991, see especially claims 1, 5, 11 and 21	1-3
<b>A</b> .	Dialog Information Services, file 5, BIOSIS 69-91, Dialog Acc.No. 7095055, Sjöberg J et al: "Insulin-like growth factor IGF-1 as a stimulator of regeneration in the freeze-injureo rat sciatic nerve", & Brain Res 485 (1), 1989, p102-108	1-4
<b>A</b>	Dialog Information Services, file 155, Medline 66-91, Dialog Acc.No. 05445532, Whitfield H.J. et al: "Isolation of a cDNA clone encoding rat isulin-like growth factor-II precursor", & Nature Nov 15-21 1984; 312 (5991) p277-80	1-4
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THE SECOND SHEET	
FURTHER INFORMATION CONTINUED FR M THE SECOND SHEET	
V. X OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE	
This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following re	asons:
1. Claim numbers.58, because they relate to subject matter not required to be searched by this Authority, namely.	
See PCT Rule 39.1(iv): Methods for treatment of the human	
or animal body by surgery or therapy, as well as diagnosti	c
methods.	
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2. Claim numbers, because they relate to parts of the international application that do not comply with the prescrib requirements to such an extent that no meaningful international search can be carried out, specifically:	_
3. Claim numbers because they are dependent claims and are not drafted in accordance with the second and third tences of PCT Rule 8.4(a).	en-
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VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING 2	
VI. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING <sup>2</sup> This International Searching Authority found multiple inventions in this international application as follows:	
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# ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO.PCT/SE 92/00275

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the Swedish Patent Office EDP file on The Swedish Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

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